

HS Prime Tag DNA Polymerase

Product Name	Cat. No.	Size
HS Prime Taq DNA Polymerase	G-7000	250 Units X 1
	G-7001	250 Units X 2
	G-7002	250 Units X 4

Package Information

G-7000	1. HS Prime Tag DNA Polymerase (250 Units, 2.5 U/ \$\mu\$, 100\$\$\mu\$ X 1)
	1. HS Prime Taq DNA Polymerase (250 Units, 2.5 U/µl , 100µl X 1) 2. 10X Reaction buffer (with 20 mM MgCl2, 1.0 ml X 1) 3. 10 mM dNTPs Mixture (2.5 mM each of dATP, dCTP, dGTP
	3. 10 mM dNTPs Mixture (2.5 mM each of dATP, dCTP, dGTP
	and dTTP. 0.5 ml X 1)

Description

HS Prime Taq DNA Polymerase is designed for hot-start PCR, a technique that enhances the specificity, sensitivity and yield of DNA amplification. In addition, the enzyme provides the convenience of reaction set-up at room temperature. The enzyme is inactived at room temperature, avoiding extension of non-specifically annealed primers or primer dimers and providing higher specificity of DNA amplification. The functional activity of the enzyme is restored during 10 minute incubation at 94°C. The activated enzyme maintains the same functionality as Taq DNA Polymerase.: it catalyzes 5'—3' synthesis of DNA, has no detectable 3'—5' proofreading exonuclease activity.

Buffers and Reagent

Storage Buffer

20 mM Tris-HCI (pH 8.0), 100 mM KCI, 0.5 mM EDTA, 0.1 mM DTT, 0.5% Tween 20, 0.5% Nonidet P-40 and 50% Glycerol

10X Reaction Buffer

Contains Tris-HCl (pH 9.0), PCR enhancers, (NH4)2SO4 and 20 mM MgCl2

10 mM dNTPs Mixture

2.5 mM each of dATP, dCTP, dGTP and dTTP

Applications

Hot-start PCR, RT-PCR, Amplification of low copy or high range size DNA targets, Real-time PCR, Multiplex PCR, T-vector cloning.

- Research Use Only
- ♦ Store at -20℃

Protocol

Optimal reaction conditions, such as reaction temperature & times, and amount of template DNA, may vary and must be individually determined.

The following 20 # reaction volume can be used for PCR.

- 1. Thaw the 10X Reaction buffer and 10 mM dNTPs Mixture.
- 2. Prepare a mastermix.

Component	Volume	Final conc.	
Sterilized D.W.	add up to 20 µl	-	
10X Reaction Buffer	2.0 <i>µ</i> l	1X	
10 mM dNTPs Mixture	0.5~2.0µl	0.25~1.0 mM	
Upstream Primer (10 pmoles/μℓ)	0.2~2.0 <i>µ</i> l	0.1~1.0 pmoles	
Downstream Primer (10 pmoles/ $\mu\ell$)	0.2~2.0 <i>µ</i> l	0.1~1.0 pmoles	
HS Prime Taq DNA Pol. (2.5 units/µl)	0.2~0.4µl	0.5~1.0 unit	
Template DNA	Variable 10 fg~1µg		

★Amount of template DNA

Bacteriophage λ , cosmid, plasmid DNA: 10 fg~300 ng

Genomic DNA: 100 ng~1µg

- 3. Mix the mastermix and dispense appropriate volumes into PCR tubes. Centrifuge the PCR tubes in a microcentrifuge for 10 seconds.
- 4. Perform PCR using your standard parameters (3-step cycling).

Step	Temp. & Time		Cycles	
Step	Temp.	Time	Cycles	
Initial denaturation	95℃	3~5 min.	1	
Denaturation	95℃	30 sec.		
Annealing	x℃	30 sec.	25~35	
Extension	72℃	30~60 sec.		
Final Extension	72℃	5 min.	1	

- ★for PCR products longer than 3~4Kb, use an extension time of approximately 1 min, per Kb DNA.
- 5. Separate the PCR products by agarose gel electrophoresis and visualize with EtBr or any other means.
- ★A DNA fragment which is amplified by HS Prime Taq DNA Polymerase has A-overhang, and it enables you to do cloning by using T-vector.